Downloaded from dmd.aspetjournals.org at ASPET Journals on June

# Cannabidiol-2',6'-Dimethyl Ether, a Cannabidiol Derivative, Is a Highly Potent and Selective 15-Lipoxygenase Inhibitor

Shuso Takeda, <sup>1</sup> Noriyuki Usami, Ikuo Yamamoto, and Kazuhito Watanabe

Organization for Frontier Research in Preventive Pharmaceutical Sciences (S.T., K.W.) and Department of Hygienic Chemistry, Faculty of Pharmaceutical Sciences (K.W.), Hokuriku University, Kanazawa, Japan; and Department of Hygienic Chemistry, School of Pharmaceutical Sciences, Kyushu University of Health and Welfare, Nobeoka, Japan (N.U., I.Y.)

Received January 26, 2009; accepted April 27, 2009

### ABSTRACT:

The inhibitory effect of nordihydroguaiaretic acid (NDGA) (a non-selective lipoxygenase (LOX) inhibitor)-mediated 15-LOX inhibition has been reported to be affected by modification of its catechol ring, such as methylation of the hydroxyl group. Cannabidiol (CBD), one of the major components of marijuana, is known to inhibit LOX activity. Based on the phenomenon observed in NDGA, we investigated whether or not methylation of CBD affects its inhibitory potential against 15-LOX, because CBD contains a resorcinol ring, which is an isomer of catechol. Although CBD inhibited 15-LOX activity with an  $IC_{50}$  value (50% inhibition concentration) of 2.56  $\mu$ M, its monomethylated and dimethylated derivatives, CBD-2′-monomethyl ether and CBD-2′,6′-dimethyl ether (CBDD), inhibited

15-LOX activity more strongly than CBD. The number of methyl groups in the resorcinol moiety of CBD (as a prototype) appears to be a key determinant for potency and selectivity in inhibition of 15-LOX. The IC $_{50}$  value of 15-LOX inhibition by CBDD is 0.28  $\mu\text{M}$ , and the inhibition selectivity for 15-LOX (i.e., the 5-LOX/15-LOX ratio of IC $_{50}$  values) is more than 700. Among LOX isoforms, 15-LOX is known to be able to oxygenate cholesterol esters in the low-density lipoprotein (LDL) particle (i.e., the formation of oxidized LDL). Thus, 15-LOX is suggested to be involved in development of atherosclerosis, and CBDD may be a useful prototype for producing medicines for atherosclerosis.

Cannabidiol (CBD) is known to be one of the major components in the cannabis plant (Mechoulam, 1970; Turner et al., 1980; Dewey, 1986; Howlett et al., 2002). It has been reported that CBD can inhibit lipoxygenase (LOX) activity with relatively high concentrations (i.e., micromolar concentration order) compared with other plant-derived inhibitors, such as luteolin, nordihydroguaiaretic acid (NDGA), and quercetin (i.e., from nanomolar to micromolar concentration order) (Evans et al., 1987; Yamamoto et al., 1998; Whitman et al., 2002; Sadik et al., 2003; Russo, 2004). In addition to the LOX inhibitors mentioned above, there are many LOX inhibitors that have been discovered in plants. LOXs are nonheme iron-containing enzymes that catalyze the dioxygenation of polyunsaturated fatty acids, such as arachidonic acid and linolenic acids. Up to now, three major LOX isoforms have been discovered (i.e., 5-, 12-, and 15-LOX) (Funk, 1996; Brash, 1999; Kühn and Thiele, 1999). Among LOX isoforms, 15-LOX is known to be able to directly oxygenate not only free fatty

This study was supported in part by the Ministry of Education, Culture, Sport, Science and Technology of Japan [Grant-in-Aid for Scientific Research (C) 20590127 (to K.W.) and Grant-in-Aid for Young Scientists (B) 20790149 (to S.T.)]; and the Academic Frontier Project for Private Universities, Ministry of Education, Culture, Sport, Science and Technology of Japan.

<sup>1</sup> Current affiliation: Department of Molecular Biology, Daiichi University, College of Pharmaceutical Sciences, Fukuoka, Japan.

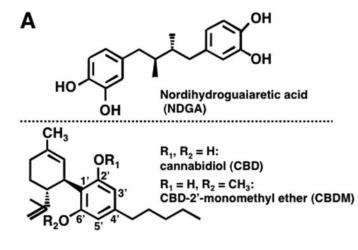
Article, publication date, and citation information can be found at http://dmd.aspetjournals.org.

doi:10.1124/dmd.109.026930.

acids but also complex substrates such as phospholipids, cholesterol ester, and the cholesterol ester in the low-density lipoprotein (LDL) particle (Brash, 1999; Kühn and Thiele, 1999; Takahashi and Yoshimoto, 2002). Oxidation of LDL is recognized as the first step for the development of atherosclerosis (Kühn et al., 1997), and the role of 15-LOX in the process of LDL oxidation and the progress of atherosclerosis has been extensively investigated using 15-LOX-knockout mice (Cyrus et al., 1999). However, the role of 15-LOX in initiating LDL oxidation is controversial, because Sparrow and Olszewski (1992) have attributed the ability of 15-LOX inhibitors to block LDL modification to nonspecific antioxidant effects rather than to direct effects on the enzyme. Thus, a 15-LOX isoform-selective inhibitor is required to clarify the involvement of 15-LOX in the formation of oxidized LDL.

However, most of the plant-derived LOX inhibitors containing catechol and resorcinol rings inhibit LOX isoforms in a nonselective manner because of their antioxidant properties, resulting in the change of cofactor Fe(III) into Fe(II) (Sadik et al., 2003). Thus, there is a need to develop highly potent and LOX isoform-selective inhibitors. It has been reported that 15-LOX activity is quite sensitive to the structural modifications of the catechol ring of NDGA, an isomer of resorcinol (Fig. 1A), such as methylation of its hydroxyl group (Blecha et al., 2007), although NDGA is well known as a pan-LOX inhibitor (Hope et al., 1983; Whitman et al., 2002). There have been no reports investigating whether any cannabinoid(s) including CBD "selectively

**ABBREVIATIONS:** CBD, cannabidiol; LOX, lipoxygenase; NDGA, nordihydroguaiaretic acid; LDL, low-density lipoprotein; CBDM, CBD-2'-mono methyl ether; CBDD, CBD-2',6'-dimehyl ether; THC,  $\Delta^9$ -tetrahydrocannabinol; CBN, cannabinol; CBDA, cannabidiolic acid; CBE, cannabielsoin; CBEM, monomethyl ether; CBDHQ, CBD-hydroxyquinone; AA-861, 2-(12-hydroxydodeca-5,10-diynyl)-3,5,6-trimethyl-1,4-benzoquinone.



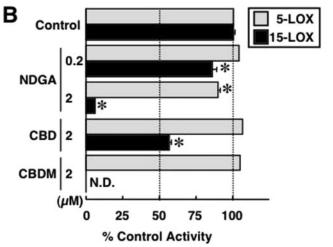


Fig. 1. Effects of CBD, CBDM, and NDGA on 5-LOX and 15-LOX activities. Structures of CBD, CBDM, and NDGA are shown in A. B, enzymatic reactions were initiated with linoleic acid, and then chromogen was added to stop the reactions and to develop colorimetric reactions. The absorbance was monitored at 490 nm. The assay conditions are described under *Materials and Methods*. Each bar represents the mean  $\pm$  S.D. (triplicate determinations) of the relative activity to the control. \*, significantly different (p < 0.05) from control. N.D., not detectable (because of complete inhibition). Numbers in CBD and CBDM correspond to the monoterpenoid nomenclature.

and potently" inhibit the 15-LOX isoform, whereas CBD having a resorcinol ring has been shown to inhibit LOX activity (Evans et al., 1987; Russo, 2004).

Based on the possibility of CBD as a prototype for a 15-LOX inhibitor, 2'-monomethylated CBD (CBDM) (Fig. 1A) and 2',6'-dimethylated CBD (CBDD) (Fig. 5A), whose respective phenolic hydroxyl groups of the resorcinol ring are methylated, were synthesized from CBD. It was observed that unlike CBD, these methylated derivatives are highly potent and selective inhibitors for 15-LOX, especially CBDD, which lacks free phenolic hydroxyl groups (IC<sub>50</sub> 280 nM), without an inhibitory effect on 5-LOX. The potential utility of CBDD as a probe for analyzing the formation of oxidized LDL is discussed.

## Materials and Methods

Cannabinoids and Chemicals.  $\Delta^9$ -Tetrahydrocannabinol (THC), cannabinol (CBN), CBD, and cannabidiolic acid (CBDA) were isolated and purified from cannabis leaves according to the methods described elsewhere (Aramaki et al., 1968). CBDM, CBDD, cannabielsoin (CBE), and CBE monomethyl ether (CBEM) were prepared as described previously (Gohda et al., 1990).

CBD-hydroxyquinone (CBDHQ) was synthesized according to the method of Mechoulam et al. (1968). Purities of these cannabinoids were determined to be at least above 95% by gas chromatography (Watanabe et al., 2005; Takeda et al., 2008). NDGA was purchased from Cayman Chemical (Ann Arbor, MI). All other reagents were of analytical grade.

Enzyme Sources. 5-LOX and 15-LOX activities were measured using a commercially available LOX inhibitor screening assay kit (Cayman Chemical). 5-LOX (lot no. 0400028-1) and 15-LOX (lot nos. 193367–193368) screening enzymes were purchased from Cayman Chemical. All inhibitors added to the reaction system were prepared just before use. After enzyme reactions, the resulting hydroperoxides were treated with chromogen to develop the reaction and then absorbance intensities were determined spectrophotometrically with a 96-well plate reader at 490 nm. No colorimetric change was observed in control incubations that were performed by omitting enzymes or with heat-denatured enzymes and inhibitors in combination with chromogen. The concentrations of cannabinoids used in this study were determined on the basis of the solubility and the concentration without interference with chromogen. Each assay was performed in triplicate.

**Data Analysis.** The concentration of the inhibitor that is required to produce 50% inhibition of the enzymatic activity (IC $_{50}$ ) was determined from the curves plotting enzymatic activity versus inhibitor concentrations using Origin7.5J software (OriginLab Corp., Northampton, MA). The details of the calculations were described in our previous articles (Takeda et al., 2006, 2008). Differences were considered to be significant when the p value was calculated to be <0.05. All statistical analyses were performed by using Scheffé's F test, which is a type of post hoc test for analyzing results of analysis of variance testing. These calculations were done using Statview5.0J software (SAS Institute Inc., Cary, NC).

#### Results

Effects of Cannabidiol and Its Derivative on LOX Activity. In the present study, we first synthesized a derivative of CBD, CBDM (Fig. 1A), because it was reported that the inhibitory effect of NDGA on 15-LOX activity was shown to be modified by methylation of the hydroxyl group of its catechol ring (Blecha et al., 2007). The inhibitory effects of CBD, CBDM, and NDGA on 5/15-LOX-catalyzed oxygenation activity were examined using purified LOX as an enzyme source. Although 5-LOX activity was not significantly inhibited by the addition of 2  $\mu$ M CBD and CBDM except for NDGA, 15-LOX activity was quite sensitive to the inhibitors investigated, particularly CBDM (complete inhibition) and NDGA (94% inhibition), compared with control activity (100%) (Fig. 1B; see also structures in Fig. 1A). It is important to know the inhibition selectivity (5-LOX/15-LOX ratio of IC<sub>50</sub> values) of CBD and CBDM-mediated inhibition of LOX isoforms. Thus, we next determined IC50 values for the inhibition of the two LOX isoforms by CBD and CBDM. It was shown that CBD had the potential to inhibit both 5- and 15-LOX isoforms, but 15-LOX activity was more selectively inhibited by CBD than by 5-LOX, which gives rise to the ratio of 28.8 (Fig. 2, A and B; Table 1). On the other hand, compared with CBD-produced inhibition of 15-LOX enzyme, CBDM exhibited a much greater inhibitory effect on 15-LOX activity than CBD did; namely, 1) the  $IC_{50}$  value of 15-LOX inhibition is 720 nM, which is approximately 3.6 times stronger than the concentration of CBD, and 2) 5-LOX activity was not remarkably inhibited even in the presence of 200  $\mu$ M CBDM (approximately 75%), although 5-LOX was inhibited by CBD with an IC $_{50}$  value of 73.73  $\mu$ M. Thus, the inhibition selectivity of CBDM was determined to be at least 278, which is approximately 9.6 times greater than that of CBD (Fig. 2, A and B; Table 1). In short, it was revealed that CBDM, which has a methylated 2'-hydroxyl group, is much more effective/selective in inhibition of 15-LOX than CBD. Based on the results obtained in Fig. 2, the following experiments focused on the inhibition of 15-LOX activity by "methylated" CBD.

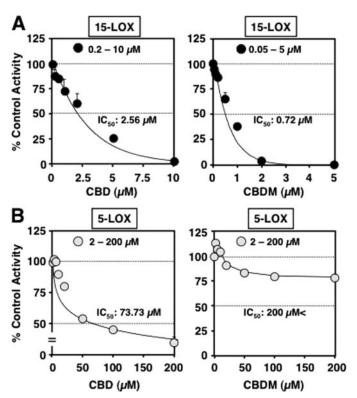


Fig. 2. Dose-dependent inhibition by CBD and CBDM on LOX activity. A and B, lipoxygenation by two isoforms of LOX enzymes (A, 15-LOX: B, 5-LOX) was examined in the presence of the indicated concentrations of CBD and CBDM. Reactions were initiated with linoleic acid, and then chromogen was added to stop the reactions and to develop colorimetric reactions. The absorbance was monitored at 490 nm. The assay conditions are described under *Materials and Methods*. Each bar represents the mean  $\pm$  S.D. (triplicate determinations) of the relative activity to the control.

TABLE 1 Comparison of  $IC_{50}$  values for CBD and its methylated forms

Inhibitors	IC <sub>50</sub> Values		
	5-LOX	15-LOX-	5-LOX/15-LOX Ratio <sup>a</sup>
	μ	иM	
CBD	73.73	2.56	28.8
CBDM	>200	0.72	278
CBDD	>200	0.28	714

 $<sup>^</sup>a$  The ratio of the IC<sub>50</sub> values for 5-LOX and 15-LOX can be used as an indication of the 15-LOX selectivity of inhibitors. A 5-LOX/15-LOX ratio of more than 1 indicates preferential 15-LOX selectivity.

Structural Requirement for Inhibitory Effect of CBD and Its **Derivatives on 15-LOX Activity.** It is well known that the inhibitory potential for LOXs by flavonoids depends on the details of their structures (e.g., the number of aromatic rings and hydroxyl groups, planarity, and so on). Here, we performed experiments to obtain information about structure-inhibition relationships by focusing on the structures of CBD and its structurally related cannabinoids (i.e., CBDA, CBE, CBEM, CBN, and THC) (Fig. 3A). 15-LOX activities were inhibited by 10  $\mu$ M CBN, 2  $\mu$ M CBD, and 0.7  $\mu$ M CBDM to the same extent ( $\sim$ 50%) compared with controls (Fig. 3B). The concentrations of CBD and CBDM were determined based on their IC<sub>50</sub> values for 15-LOX inhibition (Fig. 2; Table 1). The degree of inhibition by THC was much stronger than that of CBN, although THC was a weak inhibitor for 15-LOX compared with CBDM (Fig. 2, A and B), indicating that the free 6'-substitution group, not available for ring formation with the terpene moiety, is important for the

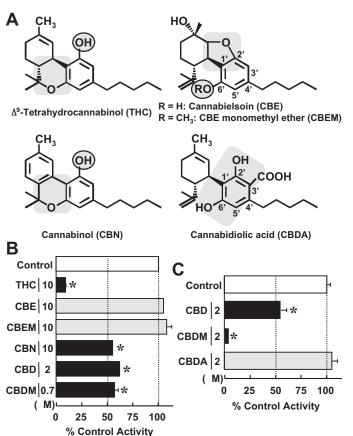


Fig. 3. Effects of CBD and its structurally related cannabinoids on 15-LOX activity. A, structures of cannabinoids tested (CBDA, CBE, CBEM, CBN, and THC). B and C, effects of CBD and its structurally related cannabinoids on 15-LOX activity were examined in the presence of the indicated concentrations of cannabinoids. Reactions were initiated with linoleic acid, and then chromogen was added to stop the reactions and to develop colorimetric reactions. The absorbance was monitored at 490 nm. The assay conditions are described under *Materials and Methods*. Each bar represents the mean  $\pm$  S.D. (triplicate determinations) of the relative activity to the control. \*, significantly different (p < 0.05) from control. Numbers in cannabinoids correspond to the monoterpenoid nomenclature.

inhibition, and the number of aromatic rings is also important. CBDA, an acid form of CBD with a carboxyl group at its 3'-position (Fig. 3A), did not have any inhibitory effect on the activity (Fig. 3C). In agreement with this result, it has been reported that 2,4-dihydroxybenzoic acid does not fit into the active site of 15-LOX (Borbulevych et al., 2004). Furthermore, CBE and CBEM, whose 2'-hydroxyl groups are used in ring formation with neighbor structures, did not exert any inhibitory effects on 15-LOX activity (Fig. 3). Taken together, these results suggested that the core resorcinol itself is not substituted and is also freed from ring formation with neighboring structures (i.e., planar structure is unfavorable), key determinants for the inhibition.

15-LOX Activity Is Not Sensitive to a Hydroxyquinone Form of CBD. This experiment was performed to investigate whether CBDHQ (Fig. 4A), an oxidized product of CBD (Watanabe et al., 1991; Usami et al., 2008), is a possible active form of CBD-mediated 15-LOX inhibition because it has been suggested that LOX enzymes have co-oxidase activity as well as oxygenase activity (Kulkarni, 2001). Furthermore, the formation of oxygenated products by 15-LOX activity has been reported to be inhibited by AA-861, a benzoquinone derivative (Li et al., 2004), although AA-861 exhibits a preference for the inhibition of 5-LOX (Ashida et al., 1983). However, 15-LOX activity was not inhibited by CBDHQ, whereas the same concentration of CBD inhibited 15-LOX activity (13%) (Fig. 4B; see also Fig.

1736 TAKEDA ET AL.

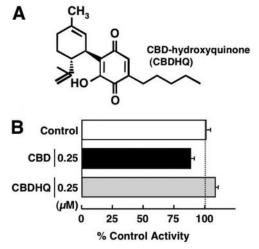


Fig. 4. A, structure of CBDHQ. B, 15-LOX activity was examined in the presence of 0.25  $\mu$ M CBDHQ or CBD (the concentration was determined based on the inhibition of CBD for 15-LOX: see Fig. 2A). Reactions were initiated with linoleic acid, and then chromogen was added to stop the reactions and to develop colorimetric reactions. The absorbance was monitored at 490 nm. The assay conditions are described under *Materials and Methods*. Each bar represents the mean  $\pm$  S.D. (triplicate determinations) of the relative activity to the control. Interference was observed when CBDHQ was added at concentrations greater than 0.5  $\mu$ M.

2A). Thus, it is suggested that CBD may exert its inhibitory effect on 15-LOX as CBD itself but not as the quinone form.

Effect of Dimethylated Form of CBD on 5/15-LOX Activity. Evidence from the present study so far suggests that methylation of the hydroxyl group of resorcinol in CBD (i.e., CBDM) was found to be effective in inhibition of 15-LOX with a reduced inhibitory potential toward 5-LOX. However, 5-LOX activity was inhibited by CBDM in a concentration-dependent manner (Fig. 2B). We hypothesized that if methylation of the hydroxyl group at the 2'-position on CBD is effective for 15-LOX inhibition, further methylation of the remaining free hydroxyl group at the 6'-position in CBDM into a methoxyl group (i.e., CBDD) may strengthen the degree of CBDMmediated inhibition of 15-LOX. Thus, we next synthesized CBDD (Fig. 5A) and subjected it to an inhibition assay against 5/15-LOX. In support of our expectation, it was revealed that CBDD was the most effective inhibitor for 15-LOX without inhibition of 5-LOX compared with CBD and CBDM (Figs. 2 and 4B; Table 1). The IC<sub>50</sub> value for 15-LOX inhibition is 0.28 µM, which has approximately 2.6- and 9.1-fold stronger potential than CBDM and CBD, respectively. The 5-LOX/15-LOX ratio of IC<sub>50</sub> values was determined to be at least 700 (Fig. 5B; Table 1). Taken together with the results obtained in Fig. 2, these results clearly demonstrated that the methylation of the resorcinol ring in CBD is highly effective in 15-LOX-selective inhibition.

# Discussion

In the current study, it was revealed that CBD and its methylated forms, CBDM and CBDD, have a selective inhibitory effect on 15-LOX-catalyzed oxygenation. In particular, CBDD was shown to be a highly potent and selective inhibitor for 15-LOX (Fig. 5B; Table 1). It was reported that quercetin is able to inhibit 15-LOX activity by competitive mechanisms, although it contains resorcinol and catechol, having antioxidant activity in the structure (Borbulevych et al., 2004). To explain this phenomenon, it was reported that quercetin is subjected to oxidation via co-oxidase activity of LOX during the oxygenation reaction. Thus, we first focused on the metabolites of CBD and CBDM as possible ultimate forms for 15-LOX inhibition because they can be metabolized into CBE and CBEM, respectively, by

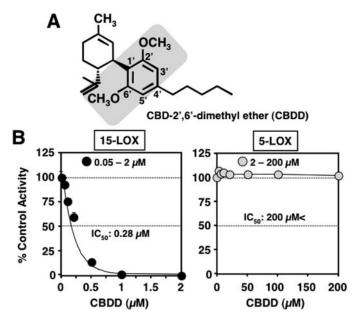


Fig. 5. Dose-dependent inhibition by CBDD on 15-LOX activity. A, structure of CBDD. B, effect of CBDD on 5-LOX and 15-LOX activity was examined in the presence of indicated concentrations of cannabinoids. Reactions were initiated with linoleic acid, and then chromogen was added to stop the reactions and to develop colorimetric reactions. Effects of CBD and CBDM on LOX activities were also investigated in this experiment, and similar results were obtained (see Fig. 2; data not shown). The absorbance was monitored at 490 nm. The assay conditions are described under *Materials and Methods*. Each bar represents the mean  $\pm$  S.D. (triplicate determinations) of the relative activity to the control.

oxidation (Fig. 3A) (Gohda et al., 1990; Yamamoto et al., 1991). However, 15-LOX activity was not inhibited even with a high concentration of CBE (10 µM) and also CBEM (10 µM) (Fig. 3B), suggesting that the cannabielsoin-type metabolites are not involved in inhibition by CBD and CBDM. CBD can also be metabolized into CBDHQ by oxidation such as cytochrome P450 (P450)-catalyzed reaction (Bornheim and Grillo, 1998). As mentioned above, LOX has co-oxidase activity as well. We next investigated whether or not CBDHQ, an oxidized product of CBD, can affect 15-LOX activity. However, 15-LOX was not inhibited by CBDHO (Fig. 4B), and no inhibition by CBDHQ was observed for 5-LOX (data not shown). We reported previously that no demethylated products were observed when CBDM and CBDD were reacted with guinea pig liver microsomes that contain many drug-metabolizing enzymes including cytochrome P450 (Gohda et al., 1990; Yamamoto et al., 1991; Oguri et al., 1994; Ishii et al., 2005; Takeda et al., 2005a,b, 2009), although CBDM can be converted into CBEM in microsomes (Gohda et al., 1990; Yamamoto et al., 1991). Thus, it is thought that the inhibition potential of CBD and CBDM might be weaken in near in vivo situations, although in the absence of drug-metabolizing enzymes, CBD and CBDM behave as selective 15-LOX inhibitors (Fig. 2; Table 1). Taking into consideration these lines of evidence, it is suggested that 15-LOX inhibition by CBDM and CBDD, including CBD, might be caused by their structural natures, although we could not exclude the possibility that CBD and CBDM-mediated inhibition of 15-LOX is also attributable to its antioxidant property (i.e., phenol and resorcinol moieties). LOX enzymes are mostly known for their peroxidase activity and for metabolizing unsaturated fatty acids. However, until now, their co-oxidase activity has been much less explored. Studies on molecular mechanism(s) underlying CBDD-mediated inhibition of 15-LOX are underway.

It has been reported that THC can suppress the progression of atherosclerosis via cannabinoid receptors in animal models, whereas CBDD was shown to have no affinity for the receptor (Thomas et al., 2004; Steffens and Mach, 2006). THC-mediated inhibition of 15-LOX is very weak compared with that of CBDD (i.e., the IC $_{50}$  values are 2.42 and 0.28  $\mu$ M, respectively) (Figs. 3B and 5B), suggesting that these cannabinoids may be able to abrogate atherosclerosis via different approaches.

Some pharmaceutical agents for the therapy of atherosclerosis can decrease LDL levels. However, the agents tend to lower total LDL, but oxidized LDL and oxidized LDL can be generated by nonspecific free radical oxidation (Sparrow and Olszewski, 1992). We propose the possibility that CBDD might be a useful prototype for producing a specific 15-LOX inhibitor without containing a phenolic hydroxyl group in its structure different from NDGA, CBD, and CBDM (Figs. 1A and 5A). Furthermore, CBDD can be used as a probe in analyzing mechanism(s) of LDL oxidation by 15-LOX because of potent and 15-LOX isoform-selective inhibition.

#### References

- Aramaki H, Tomiyasu N, Yoshimura H, and Tsukamoto H (1968) Forensic chemical study on marihuana. I. A detection method of the principal constituents by thin-layer and gas chromatographies. Chem Pharm Bull 16:822–826.
- Ashida Y, Saijo T, Kuriki H, Makino H, Terao S, and Maki Y (1983) Pharmacological profile of AA-861, a 5-lipoxygenase inhibitor. *Prostaglandins* 26:955–972.
- Blecha JE, Anderson MO, Chow JM, Guevarra CC, Pender C, Penaranda C, Zavodovskaya M, Youngren JF, and Berkman CE (2007) Inhibition of IGF-1R and lipoxygenase by nordihydroguaiaretic acid (NDGA) analogs. *Bioorg Med Chem Lett* 17:4026–4029.
- Borbulevych OY, Jankun J, Selman SH, and Skrzypczak-Jankun E (2004) Lipoxygenase interactions with natural flavonoid, quercetin, reveal a complex with protocatechuic acid in its X-ray structure at 2.1 Å resolution. *Proteins* 54:13–19.
- Bornheim LM and Grillo MP (1998) Characterization of cytochrome P450 3A inactivation by cannabidiol: possible involvement of cannabidiol-hydroxyquinone as a P450 inactivator. Chem Res Toxicol 11:1209–1216.
- Brash AR (1999) Lipoxygenases: occurrence, functions, catalysis, and acquisition of substrate. *J Biol Chem* **274**:23679–23682.
- Cyrus T, Witztum JL, Rader DJ, Tangirala R, Fazio S, Linton MF, and Funk CD (1999) Disruption of the 12/15-lipoxygenase gene diminishes atherosclerosis in apo E-deficient mice. J Clin Invest 103:1597–1604.
- Dewey WL (1986) Cannabinoid pharmacology. Pharmacol Rev 38:151-178.
- Evans AT, Formukong EA, and Evans FJ (1987) Actions of cannabis constituents on enzymes of arachidonate metabolism: anti-inflammatory potential. *Biochem Pharmacol* 36:2035–2037.
- Funk CD (1996) The molecular biology of mammalian lipoxygenases and the quest for eico-sanoid functions using lipoxygenase-deficient mice. *Biochim Biophys Acta* 1304:65–84.
- Gohda H, Narimatsu S, Yamamoto I, and Yoshimura H (1990) In vivo and in vitro metabolism of cannabidiol monomethyl ether and cannabidiol dimethyl ether in the guinea pig: on the formation mechanism of cannabielsoin-type metabolite from cannabidiol. *Chem Pharm Bull* 38:1697–1701.
- Hope WC, Welton AF, Fiedler-Nagy C, Batula-Bernardo C, and Coffey JW (1983) In vitro inhibition of the biosynthesis of slow reacting substance of anaphylaxis (SRS-A) and lipoxygenase activity by quercetin. *Biochem Pharmacol* 32:367–371.
- Howlett AC, Barth F, Bonner TI, Cabral G, Casellas P, Devane WA, Felder CC, Herkenham M, Mackie K, Martin BR, et al. (2002) International Union of Pharmacology. XXVII. Classification of cannabinoid receptors. *Pharmacol Rev* 54:161–202.
- Ishii Y, Takeda S, Yamada H, and Oguri K (2005) Functional protein-protein interaction of drug metabolizing enzymes. Front Biosci 10:887–895.
- Kühn H, Heydeck D, Hugou I, and Gniwotta C (1997) In vivo action of 15-lipoxygenase in early stages of human atherogenesis. J Clin Invest 99:888–893.
- Kuhn H and Thiele BJ (1999) The diversity of the lipoxygenase family: many sequence data but little information on biological significance. FEBS Lett 449:7–11.

- Kulkarni AP (2001) Lipoxygenase—a versatile biocatalyst for biotransformation of endobiotics and xenobiotics. Cell Mol Life Sci 58:1805–1825.
- Li Q, Cheon YP, Kannan A, Shanker S, Bagchi IC, and Bagchi MK (2004) A novel pathway involving progesterone receptor, 12/15-lipoxygenase-derived eicosanoids, and peroxisome proliferator-activated receptor gamma regulates implantation in mice. *J Biol Chem* 279: 11570–11581.
- Mechoulam R (1970) Marihuana chemistry. Science 168:1159-1166.
- Mechoulam R, Ben-Zvi Z, and Gaoni Y (1968) Hashish. 13. On the nature of the Beam test. Tetrahedron 24:5615–5624.
- Oguri K, Yamada H, and Yoshimura H (1994) Regiochemistry of cytochrome P450 isozymes. Annu Rev Pharmacol Toxicol 34:251–279.
- Russo EB (2004) Clinical endocannabinoid deficiency (CECD): can this concept explain therapeutic benefits of cannabis in migraine, fibromyalgia, irritable bowel syndrome and other treatment-resistant conditions? *Neuro Endocrinol Lett* 25:31–39.
- Sadik CD, Sies H, and Schewe T (2003) Inhibition of 15-lipoxygenases by flavonoids: structure-activity relations and mode of action. Biochem Pharmacol 65:773–781.
- Sparrow CP and Olszewski J (1992) Cellular oxidative modification of low density lipoprotein does not require lipoxygenases. *Proc Natl Acad Sci U S A* **89:**128–131.
- Steffens S and Mach F (2006) Cannabinoid receptors in atherosclerosis. Curr Opin Lipidol 17:519–526.
- Takahashi Y and Yoshimoto T (2002) What are the functions of mammalian 8-, 12-, and 15-lipoxygenases? Res Adv Cancer 2:221-229.
- Takeda S, Ishii Y, Iwanaga M, Mackenzie PI, Nagata K, Yamazoe Y, Oguri K, and Yamada H (2005a) Modulation of UDP-glucuronosyltransferase function by cytochrome P450: evidence for the alteration of UGT2B7-catalyzed glucuronidation of morphine by CYP3A4. Mol Pharmacol 67:665–672.
- Takeda S, Ishii Y, Mackenzie PI, Nagata K, Yamazoe Y, Oguri K, and Yamada H (2005b) Modulation of UDP-glucuronosyltransferase 2B7 function by cytochrome P450s in vitro: differential effects of CYP1A2, CYP2C9 and CYP3A4. Biol Pharm Bull 28:2026–2027.
- Takeda S, Ishii Y, Iwanaga M, Nurrochmad A, Ito Y, Mackenzie PI, Nagata K, Yamazoe Y, Oguri K, and Yamada H (2009) Interaction of cytochrome P450 3A4 and UDP-glucuronosyltransferase 2B7: evidence for protein-protein association and possible involvement of CYP3A4 J-helix in the interaction. Mol Pharmacol 75:956–964.
- Takeda S, Kitajima Y, Ishii Y, Nishimura Y, Mackenzie PI, Oguri K, and Yamada H (2006) Inhibition of UDP-glucuronosyltransferase 2B7-catalyzed morphine glucuronidation by ketoconazole: dual mechanisms involving a novel noncompetitive mode. *Drug Metab Dispos* 34:1277-1282.
- Takeda S, Misawa K, Yamamoto I, and Watanabe K (2008) Cannabidiolic acid as a selective cyclooxygenase-2 inhibitory component in cannabis. *Drug Metab Dispos* 36:1917–1921.
- Thomas A, Ross RA, Saha B, Mahadevan A, Razdan RK, and Pertwee RG (2004) 6"-Azidohex-2"-yne-cannabidiol: a potential neutral, competitive cannabinoid CB1 receptor antagonist. Eur J Pharmacol 487:213–221.
- Turner CE, Elsohly MA, and Boeren EG (1980) Constituents of Cannabis sativa L. XVII. A review of the natural constituents. J Nat Prod 43:169–234.
- Usami N, Yamamoto I, and Watanabe K (2008) Generation of reactive oxygen species during mouse hepatic microsomal metabolism of cannabidiol and cannabidiol hydroxy-quinone. *Life Sci* 83:717–724.
- Watanabe K, Motoya E, Matsuzawa N, Funahashi T, Kimura T, Matsunaga T, Arizono K, and Yamamoto I (2005) Marijuana extracts possess the effects like the endocrine disrupting chemicals. *Toxicology* 206:471–478.
- Watanabe K, Usami N, Yamamoto I, and Yoshimura H (1991) Inhibitory effect of cannabidiol hydroxy-quinone, an oxidative product of cannabidiol, on the hepatic microsomal drugmetabolizing enzymes of mice. J Pharmacobiodyn 14:421–427.
- Whitman S, Gezginci M, Timmermann BN, and Holman TR (2002) Structure-activity relationship studies of nordihydroguaiaretic acid inhibitors toward soybean, 12-human, and 15-human lipoxygenase. J Med Chem 45:2659–2661.
- Yamamoto I, Gohda H, Narimatsu S, Watanabe K, and Yoshimura H (1991) Cannabielsoin as a new metabolite of cannabidiol in mammals. *Pharmacol Biochem Behav* 40:541–546.
- Yamamoto H, Sakakibara J, Nagatsu A, and Sekiya K (1998) Inhibitors of arachidonate lipoxygenase from defatted perilla seed. J Agric Food Chem 46:862–865.

Address correspondence to: Dr. Kazuhito Watanabe, Department of Hygienic Chemistry, Faculty of Pharmaceutical Sciences, Hokuriku University, Ho-3 Kanagawa-machi, Kanazawa 920-1181, Japan. E-mail: k-watanabe@hokuriku-u. ac.jp